

Infection - pain in oral cavity and alternative managements

PhD thesis

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List of Abbreviations

CD	Cluster of differentiation
CGRP	Calcitonin gene-related peptide
CI	Confidence interval
DRG	Dorsal root ganglia
GABA	Gamma aminobutyric acid
GRADE	Grading of Recommendations, Assessment, Development and Evaluation
FPR	Formyl peptide receptor
HHV	Human herpes virus
HSV	Herpes simplex virus
HZI	Herpes zoster infection
HZV	Herpes zoster virus
IASP	The International Association for the Study of Pain
IL	Interleukin
LPS	Lipopolysaccharide
MAPK	Mitogen-activated protein kinase
OR	Odds Ratio
PAMP	Pathogen-associated molecular pattern
PHN	postherpetic neuralgia
PRR	Pattern recognition receptor
PICO	Population, intervention, control, outcome
PRISMA	Preferred Reporting Items for Systematic Reviews and Meta-Analyses
RCT	Randomized clinical trial
SMD	Standardized mean difference
SP	Substance P
TLR	Toll-like receptor
TRP	Transient receptor potential channel
TRPA	Transient Receptor Potential Channel, Ankyrin subtype
TRPV	Transient Receptor Potential Channel, Vanilloid subtype
VZV	Varicella zoster virus

1. Introduction

1.1. Pain

Pain is a complex perception involving biological, psychological, and social factors. The International Association for the Study of Pain (IASP) has revised and defined pain as “an unpleasant sensory and emotional experience associated with, or resembling that associated with, actual or potential tissue damage” (1). Normal pain sensation or nociceptive pain is evoked by stimulus which is intense enough to trigger nociceptors (Figure 1a). Primary nociceptive afferent neurons are responsible for nociceptive pain by transmitting noxious stimuli to neurons in the spinal cord and higher center (2). The sensation felt corresponds in location, times, and quality of stimulus in expected manner (3).

Tissue injuries and/or infection cause pain through inflammatory process, serving as a warning signal to individual (4). Triggered innate immune cells secrete a various type of cytokines responsible for cell-cell interaction, immune response activation, and the maintaining of cellular function (5, 6). Chemokines, cytokines and tissue specific growth factors are upregulated in the inflammatory process (7, 8). Interleukins (ILs) are the main cytokines that play important roles on immunomodulation and the development and growth of hematopoietic tissues (9). While most ILs are pro-inflammatory mediators, some ILs such as IL-4, -10, -13 show anti-inflammatory effects (10, 11).

Secreted cytokines from activated immune cells can decrease the action potential threshold of nociceptive neurons (12, 13). The action of cytokines occurs via mitogen-activated protein kinase (MAPK) and other signaling molecules, leading to the phosphorylation and gating of ion channels such as voltage-gated sodium channels and transient receptor potential (TRP) channels (14-16). Histamine, bradykinin, prostaglandin and leukotriene secreted from immune cells also have the ability to sensitize nociceptive neurons and initiate inflammatory pain (Figure 1b). The sensitized nerve becomes more sensitive to stimulation, resulting in the secretion of neuropeptides, such as calcitonin gene-related peptide (CGRP) and substance P (SP) (Figure 1c). These secreted neuropeptides act on both immune cells and nerve endings to facilitate nerve sensitization (17).

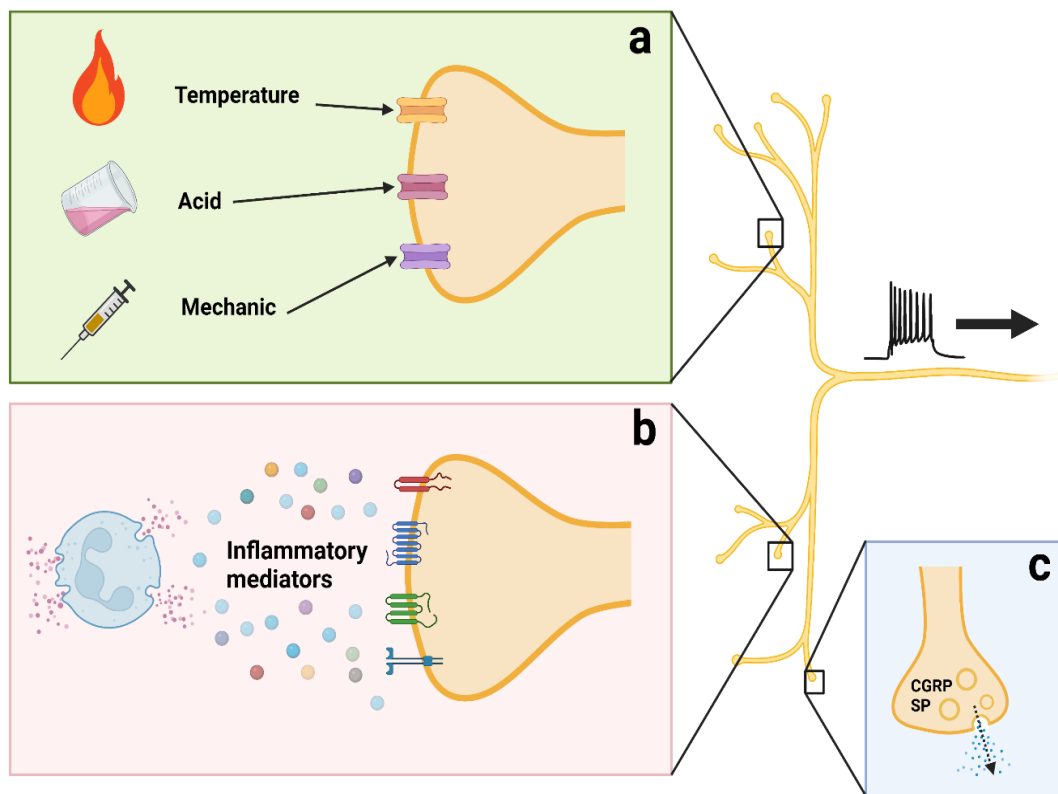


Figure 1. Nociceptive, inflammatory and neuropathic pain.

(a) Noxious stimuli are transduced into electrical activity at the peripheral terminals of unmyelinated C-fiber and thinly myelinated Aδ-fiber nociceptors by specific receptors or ion channels. The resulting input is conducted to the spinal cord and higher centers. (b) Activated immune cells release chemical mediators creating an 'inflammatory soup' that activates or modifies the stimulus response properties of nociceptor afferents. (c) The antidromic signal of the sensitized nerve fiber secretes neuropeptides at the nerve ending. Adapted from Scholz and Woolf (4), and Basbaum *et al.* (18). Created with BioRender.com.

A new aspect of pain sensation has been introduced since a discovery of pathogen recognition receptor (PRR) expression on nociceptor neurons. Nociceptive and pruriceptive neurons exhibit specific PRRs, including toll-like receptors (TLRs), formyl peptide receptors (FPRs), and signaling pathways that can sensitize transient receptor potential (TRP) channels. When microbial pathogenic ligands bind to those receptors, the nerve impulse is readily generated (Figure 2). The impulse is both orthodromically conducted to second order neurons and antidromically conducted to other axon terminals,

leading to a release of neuropeptides. Consequently, the immune system is activated and modulated. This microbe-neuron-immune interplay has been implicated in host defense mechanism (19).

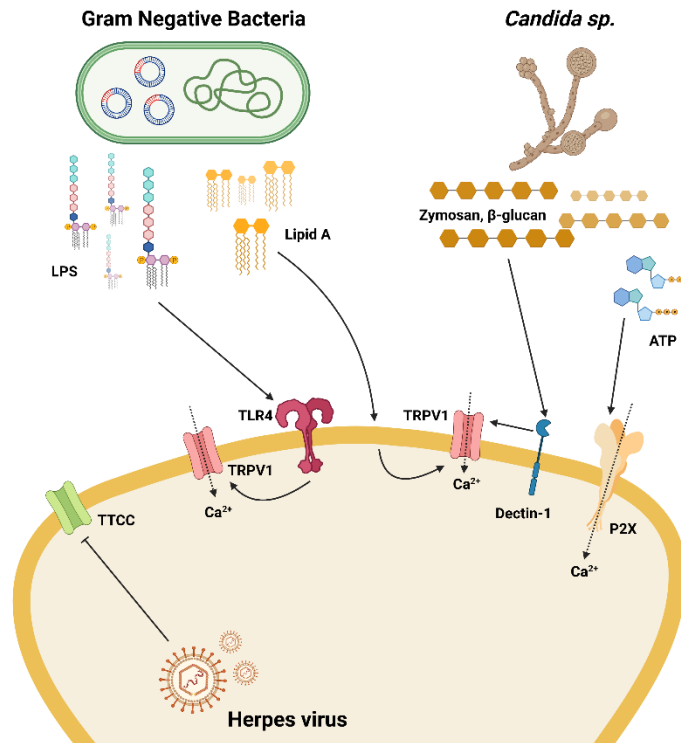


Figure 2. Molecular mechanism of microbial modulating nociception.

Bacterial, fungal and viral pathogens activate nociceptive neurons via different mechanisms. LPS: Lipopolysaccharide, ATP: Adenosine triphosphate, TTCC: T-type calcium channel, TRPV1: Transient Receptor Potential Channel, Vanilloid subtype 1, TLR4: Toll-like receptor 4. Adapted from Deng L & Chiu IM (2021) (20). Created with BioRender.com.

1.2. Oral infection

An infection is an invasion of pathogenic bacteria, viruses, fungi or other microorganisms into the body. It initiates the body reaction called inflammation, leading to swelling, redness, heat, pain and loss of function at the inflammatory site (21).

The oral cavity harbors more than 700 species of microbes, including both non-pathogenic and pathogenic microbes (22). Different ecological conditions of oral cavity (e.g. the teeth, gingiva, buccal mucosa, tongue) are colonized by distinct microbial communities. In health, there is a commensal stage between microbe and host. However,

when an imbalance between microbe-microbe or microbe-immune occurs, it can lead to oral diseases (23). For example, an overwhelming of pathogenic bacteria over non-pathogenic bacteria in oral cavity can cause periodontal diseases. The immune system intolerance contributes to oral candidiasis susceptibility, and reactivation of herpes zoster virus (HZV). Besides, interactions of pathogens with the nerve system, may result in nerve sensitization. Some pathogenic bacteria, viruses and fungi that cause diseases in the orofacial area have abilities to sensitize and activate nociceptors directly (Figure 2). Table 1 shows an example of pathogens associated with orofacial diseases that can sensitize nociceptors.

Table 1. Pathogens associated with orofacial diseases that can sensitize nociceptors.

Pathogen name	Disease	Infection site(s)	Neuronal sensitization mechanisms
Bacteria			
<i>Porphyromonas gingivalis</i>	Periodontal disease	Oral cavity	LPS sensitizes TRPV1
Virus			
Herpes simplex virus	Herpes labialis/gingivostomatitis	Trigeminal ganglia	Nociceptor sensitization
Varicella zoster virus	Orofacial herpes zoster	Trigeminal ganglia	PHN
Fungus			
<i>Candida albicans</i>	Candidiasis	Skin, oral cavity	Zymosan sensitizes nociceptor neurons

LPS: lipopolysaccharide, PHN: postherpetic neuralgia, TRPV1: transient receptor potential channel, vanilloid subtype 1. Adapted from Chui (19) with permission.

1.3. Periodontal diseases

Periodontal diseases encompass a wide variety of chronic inflammation of gingiva, bone and tooth supporting structures, initiated by bacterial dysbiosis. It begins with a localized inflammation of the gingiva – called gingivitis. An untreated gingivitis can progressively turn into periodontitis, which is characterized by the loss of the

periodontium and subsequently loss of teeth (24). Bacteria associated with periodontal diseases are dominantly Gram-negative bacteria, such as *Porphyromonas gingivalis*, *Treponema denticola*, *Tannerella forsythia*, *Prevotella intermedia*, and *Fusobacterium nucleatum* (25-28). In addition, *Aggregatibacter actinomycetemcomitans* is considered as a key pathogen of early-onset periodontitis (29).

The prevalence and incidence of periodontitis are globally estimated to be 10.8% and 701 cases per 100,000 person-year, respectively (30). Generally, individuals with periodontal diseases have been found in a higher proportion in developing and developed countries (31). Host and environmental factors also contribute to disease progression and inflammation (31).

Due to the chronic nature of periodontal diseases, clinical manifestations range from mild to severe inflammation. Initial symptoms are halitosis, gingival swelling and bleeding. This is followed by uncontrolled and exaggerated inflammation, leading to destruction of periodontal tissues. Consequently, gingival recession, bone resorption, tooth mobility and tooth loss can occur (25, 27). Typically, Periodontal diseases can cause mild-to-moderate, episodic or persistent dull pain due to infection and inflammation (32). A study on chief complaints of patients seeking for periodontitis treatment reported that 21.5% of patients who suffered from periodontitis complained about their painful gingiva and teeth (33).

During invasion and infection of pathogenic bacteria, the host may perceive pain from tissue damages and inflammatory processes, or from direct bacterial stimulation, or both. A recent review has suggested that nociceptor neurons can detect Gram-positive and Gram-negative bacterial infection through specific pathogen-recognition pathways including TLRs, FPR1, TRPA1, and TRPV1 ion channels (34).

Lipopolysaccharide (LPS) is a bacterial endotoxin found in all Gram-negative bacteria. It also serves as a pathogen-associated molecular pattern (PAMP) that is detected during infection via TLR4, CD14, and Myeloid differentiation factor 2. TRPV1-positive trigeminal neurons have been found to express TLR4 and CD14, allowing them to directly sense LPS (35). A study showed that application of LPS derived from *P. gingivalis*, a bacterium commonly found in periodontal diseases and endodontic infection, activated neurons and sensitized TRPV1 via TLR4 (36). Furthermore, LPS sensitizes neurons, resulting in Ca^{2+} influx and inward currents after capsaicin stimulation, via TRPA1 ion

channel (37). Consequently, CGRP neuropeptide is released from nerve ending (36, 38). Therefore, LPS from oral bacteria can sensitize and produce nociceptive sensation in intraoral organs, especially dental pulp and gingiva (Figure 2).

1.4. Orofacial herpes zoster

A number of viruses have been identified in the oral cavity. Some viruses have a clear role in developing oral lesions, such as oral ulcers from herpesviruses, oral warts and tumors from papillomaviruses (39). The human herpesvirus (HHV) family is a common cause of viral disease in the orofacial region. Three members of the human herpesviruses, consisting of herpes simplex virus 1 (HSV1; HHV1), herpes simplex virus 2 (HSV2; HHV2), and varicella zoster virus (VZV; HHV3), cause painful ulceration of the skin and oral mucosa. Additionally, herpesviruses can be found in periodontal pockets in associated with *P. gingivalis*, *P. intermedia*, and *T. forsythia* (40).

VZV causes varicella (chicken pox) in the primary infection. The virus then becomes latent in ganglionic neurons and glial cells (41-43). Reactivation of latent VZV infection causes herpes zoster (shingles). The viral reactivation becomes more frequent in elders who have diminished cell-mediated immunity to the virus. Moreover, immunocompromised patients are at risk for more severe diseases, such as disseminated herpes zoster (dissemination of virus in the whole body), and have a higher incidence of complications (42, 44). The infection causes damages to the infected neurons, nerve fibers, and target tissues which innervated by those infected nerves (45, 46). Consequently, the structures and functions of both peripheral and central nervous system alter, including loss of neurons and glial cells.

VZV infection frequently occurs in advanced age patients because of decreased immunity to the virus (42). The incidence of varicella ranges from 13 to 16 cases per 1,000 person-years, while the population based incidence of zoster is around 2 to 5 per 1,000 patients-years of observation (47). The varicella vaccine dramatically reduces the per year prevalence of disease (41).

Clinical manifestations of VZV infection are typically painful unilateral erythematous maculopapular rashes distributed in specific dermatomes, except for the primary infection. The rash evolves into groups of clear vesicles and subsequently ruptures, resulting in painful ulcers. Pain, hyperesthesia, and allodynia are the most common complaint of the patient (48, 49). Herpes zoster-associated pain has been

categorized as follows: acute zoster pain (acute herpetic neuralgia) which occurs after the initial rash and no longer than a month; subacute zoster pain (subacute herpetic neuralgia) which occurs between 1 to 4 month; and postherpetic neuralgia (PHN) which develops beyond 4 month since the initial onset of the rash (50). Of these, PHN is complicated and intractable herpes zoster-associated pain (50).

Generally, pain from herpes virus infection is caused by induced peripheral and central sensitization, and abnormal reorganization of nociceptive sensation (45, 46). Interesting evidence showed that the VZV cause excitation of rat DRG sensory neurons and the DRG neurons infected with VZV became more sensitive to adrenergic stimulation (51). Nociceptive neurons have been found to express TLR3 and TLR7 which recognize the single-strand RNA of viruses (52). The study of Park *et al.* found that extracellular microRNAs can stimulate DRG to fire (52). The HSV-1 infected neuron-like cell line reducing the expression of calcium channels, thereby altering electrical excitability and affecting sensory transmission (20) (Figure 2). Although the exact mechanism on how herpes viruses affect nociceptive neurons during infection and reactivation is not well understood, herpes-associated pain is previously thought to be mediated by peripheral and central neuronal changes.

Gamma aminobutyric acid (GABA)-mediated inhibition, GABA synthesis, μ -opioid receptors are destroyed from the infection, leading to the impairment of inhibitory mechanisms. Surviving neurons also demonstrate altered protein expression and phenotypic switching (45, 46). Moreover, demyelination typically occurs and lead to ephaptic crosstalk, the non-synaptic interaction between two or more demyelinated membranes. Remodeling of membranes by alteration of transducers, receptors, and ion channels can be found in herpes infection, resulting in spontaneous ectopic discharges, and hyperexcitability electrogenesis of neurons (53).

1.5. Managements of periodontal diseases

The aims of periodontal treatments are to reduce the periodontal pathogenic bacteria and to remove infected tissues, thereby provoking periodontal tissue healing (54-56). Main conventional management of periodontal disease is removing bacterial plaque and calculus by scaling and root planning. The principle of this treatment is to mechanically remove biofilm, remove supra-and infra-gingival calculus, and infected

tissue. The improvement of oral hygiene is also important to achieve and balance oral microbial symbiosis (57). However, some patients do not respond well to the conventional treatment. Therefore, several adjuvant treatments have been used, including antibiotics (24, 58), probiotics (59, 60) and photodynamic therapy (61).

Probiotics are live microorganisms, which confer a health benefit on the host when administered in adequate amounts (62). The mechanism of action of probiotics on pathogenic bacteria occurs in two modes: direct and indirect interactions (23). As the direct mechanism, probiotic microbes compete and disrupt colonization of pathogens. They can also compete for nutrients and tissue binding site, and therefore disrupt the biofilm formation. Another important and distinguished mechanism of probiotics is to produce antimicrobial substances and produce unsuitable environments for competitor species. Depending on the strain of the microbes in probiotics, different substances such as lactic acid, organic acid, peroxide, bacteriocin and anti-adhesive molecule have been reported (23, 59). The indirect mechanism of probiotics against pathogen is to modulate immune cells and alter cytokine production (23).

Since probiotics exert beneficial effects against pathogenic bacteria, some clinical trials have reported the usage of probiotics in oral health, including dental caries, halitosis, and periodontal disease (63, 64). However, the advantages of probiotics in oral health care remains unclear. One systematic review suggested that probiotics could decrease the colony forming unit of bacterial pathogens (65). On the contrary, other studies have argued that probiotic treatments do not significantly alter the pathogenic flora in the oral cavity (66-68). Recent reviews and meta-analyses on the efficacy of probiotics in periodontal diseases did not confirm the effectiveness of probiotics in decreasing the number of pathogens (69-72). Therefore, the clinical question has been addressed whether probiotics have abilities to decrease pathogenic bacteria in patients with periodontal diseases.

1.6. Management of orofacial herpes zoster

VZV are neurotropic viruses that can damage the nervous system peripherally and centrally, resulting in neuropathic pain. The goal of treatment is to control the infection and pain by using antiviral agents and analgesics (48). Antiviral agents reduce viral load by blocking viral replication, leading to a reduction in the severity and duration of

infection, and reducing the intensity of zoster pain (48). An anti-inflammatory effect from corticosteroid drugs can also reduce inflammatory pain in acute herpes zoster (73-75). As the infection causes structural and functional changes of the infected neurons, management may become more difficult. Currently available medications only treat symptoms, but not the underlying cause of the pain, by acting on membranes and synapses to reduce the degree of excitation and ectopic discharge (41).

On the basis of a systematic review and meta-analysis, the Special Interest Groups on Neuropathic Pain recommended use of gabapentinoids, Tricyclic antidepressants, and Serotonin-norepinephrine reuptake inhibitors as the first line drugs for neuropathic pain (76). Lidocaine patches and capsaicin patches are the second line drugs which can be used for peripheral neuropathic pain, while botulinum and strong opioids are the third line drugs with a weak recommendation (76). In addition to pharmacotherapy, local anesthesia, neurolytic block of sympathetic nerves, acupuncture, spinal intrathecal injection, intercostal nerve block, spinal cord stimulation, and cryotherapy have been used (77-79).

Gabapentinoids (gabapentin and pregabalin) are derivatives of GABA which binds to the $\alpha 2\text{-}\delta$ sub-unit of voltage-dependent calcium channels (80). Both substances have a similar pharmacological profile (81, 82). Gabapentinoids have an ability to inhibit ectopic discharges from peripheral nerve injuries, suppress neuralgia and sensitization, and modulate GABAergic, glutaminergic and monoaminergic functions (73, 83, 84). It has been used in patients with acute zoster infection in order to effectively control acute pain and prevent developing of chronic pain (85). Gabapentinoids may be successful in preventing postherpetic neuralgia in a long run by reducing the occurrence of acute herpetic neuralgia after a healed infection. The available literature on this effective of gabapentinoids is still inhomogeneous and has low statistical power. Therefore, the clinical question has been addressed to find the conclusion and increase statistical power on the efficacy of gabapentinoids in reducing the occurrence of acute herpes zoster pain.

2. Objectives

1. To investigate the efficacy of orally administered probiotics in reducing the number of periodontal pathogenic bacteria.
2. To investigate the effectiveness of gabapentinoids in reducing the occurrence of acute herpes zoster pain after herpes zoster infection.

In order to draw a conclusion from different available results, all objectives were investigated using meta-analysis.

3. Results

3.1. The efficacy of orally-administered probiotics to reduce the quantity of pathogenic periodontal bacteria

3.1.1. Protocol and registration

The protocol of this study was registered in the International Perspective Register of Systematic Review (PROSPERO) in the registration number CRD42018094903

3.1.2. Clinical question

The PICO framework was constructed according to the interested clinical question whether the orally administered probiotics decreases the level of pathogenic periodontal bacteria in saliva and dental plaque. The population (P) consists of patients with periodontal diseases; the intervention (I) is orally administered probiotics; the control (C) is placebo treatment or no treatment; and the outcome (O) is the amount of pathogenic periodontal bacteria in saliva, supra- and sub-gingival plaque. A systemic searching for both objectives were performed in databases. The keywords used for the search was [*probiotic AND ("periodontal disease" OR periodontitis OR gingivitis or plaque or saliva)*]

3.1.3. The result of literature search and the selection of studies

The comprehensive literature search from different databases, namely PubMed, Cochrane Central Register of Controlled Trials (CENTRAL), EMBASE, and Web of Science, yielded 2,210 records. No additional record was found through the bibliographic references of review articles. After removing duplicates, there were 1,281 records remaining and they were screened for titles and abstracts. Twenty-five remaining records were assessed for eligibility. Nine articles were excluded due to non-randomized controlled trials (86-90), non-periodontal disease participants (91, 92), sub-gingivally administration (93), and non-living bacteria (94). A further two studies were excluded because they did not fit the aim of this meta-analysis from measuring total bacterial numbers and obligate anaerobes (64, 95). Finally, fourteen articles were included in the qualitative analysis, and out of these, nine were suitable for quantitative analysis (Figure 3).

From fourteen included for qualitative analysis, five studies were excluded because of no periodontal pathogen number obtained after contacting the authors (96), unspecified participants with periodontal disease (97), used combinations of probiotics

and antibiotics (98, 99), and delay of probiotics administration (100). The remaining nine articles (63, 68, 101-107) were included for quantitative analysis.

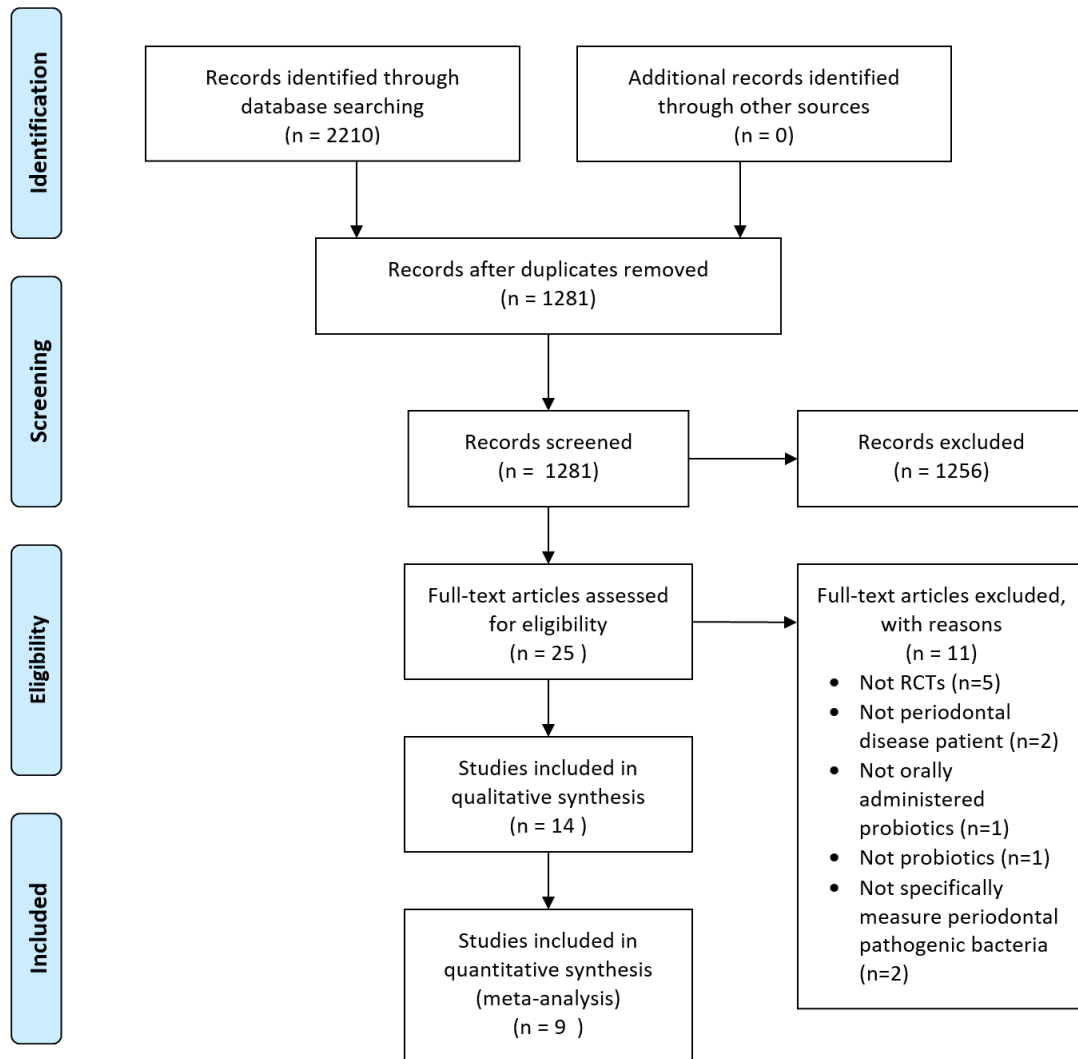


Figure 3. PRISMA 2009 flow diagram for identification of relevant studies (108).

3.1.4. Characteristics of included studies

All of the included studies are RCTs, but they differed in details. Four studies were open, controlled, parallel RCTs (97-99, 106); eight investigations were double-blinded RCTs (63, 68, 96, 101-104, 107); one study was double-blinded, crossover RCT (105), and the last one was double-blinded, split-mouth RCT (100). Each study inherited its unique in experimental design, such as different probiotic strains, forms of probiotics, instruction of use, duration of study, and the measurement of different periodontal pathogenic bacteria. The major differences among included studies for quantitative

analysis are pre-treatment and microbiologic method. Majority of studies provided scaling and root planning to the participants before starting probiotics (63, 68, 101, 102, 104, 106, 107), while two studies did not provide it (103, 105). Three studies used a conventional microbiological cultivation for measuring amount of bacteria of interest (105-107), and the rest performed a molecular method using qPCR (63, 68, 101-104, 107). The summary characteristics of the included article can be found in the publication of Sang-Ngoen *et al* (108).

	Random sequence generation (selection bias)	Allocation concealment (selection bias)	Blinding of participants and personnel (performance bias)	Blinding of outcome assessment (detection bias)	Incomplete outcome data (attrition bias)	Selective reporting (reporting bias)
Alanzi et al. 2018	+	+	+	+	+	+
Dhaliwal et al. 2017	+	-	-	-	-	+
Goyal et al. 2019	?	-	-	-	-	+
Iniesta et al. 2012	+	+	+	+	+	+
Invernici et al. 2018	+	-	+	+	+	+
Laleman et al. 2015	+	+	+	+	+	-
Laleman et al. 2019	+	+	+	+	+	+
Mayanagi et al. 2009	+	+	+	+	+	+
Montero et al. 2017	+	+	+	+	-	-
Morales et al. 2018	+	+	?	+	-	-
Shah et al. 2013	+	-	-	-	+	+
Shah et al. 2017	?	-	-	-	+	+
Teughels et al. 2013	+	+	+	+	+	+
Vivekananda et al. 2010	+	+	+	+	+	+

Figure 4. A summary of the risk of bias of included studies (108).

3.1.5. The risk of bias assessment

All included studies detailed the method of randomization and they were rated as a low risk of bias, except, one study that did not clearly specified the method (97). Thus, it was rated as having a questionable risk of bias. Five studies were determined as high risk of bias in the allocation of concealment domain because of no blinding of staffs (97-99, 104, 106). The performance bias domain was determined as high risk in four studies due to lack of blinding participants and involved personnel (97-99, 106), and questionable

risk in one study because of a chance of unblinded personnel (107). The attrition bias domain which assesses the incomplete outcome data was rated as having a high risk for four studies due to an incomplete report without further explanation (68, 97, 106, 107). Selective reporting bias from three studies were rate as having a high risk of bias because they did not report all prespecified outcomes (68, 101, 107). The results of risk of bias assessment are summarized and detailed in Figure 4 (above).

3.1.6. The result of the meta-analysis

A. actinomycetemcomitans, *P. gingivalis*, *P. intermedia*, *F. nucleatum*, and *T. forsythia* were measured after probiotics treatment in different designated follow-up duration (four-, eight-, or twelve-week). Only the *A. actinomycetemcomitans* showed the significantly positive effect of number reduction after four week of probiotics treatment, disregarding locations of samples collected (SMD= -0.28; 95%CI= -0.56 to -0.01; $p=0.045$; heterogeneity: $I^2=36.5\%$, $p=0.150$). However, the saliva, sub- or supra-gingival plaque of *A. actinomycetemcomitans* at four week did not show a significant difference between the probiotics treated group and control group (Figure 5). In addition, the probiotics treated group was not significantly different from the control group in overall result, saliva, sub-or supra-gingival plaque at eight week (Figure 6). With disregarding or regarding the locations of samples collected, the number of *P. gingivalis*, *P. intermedia*, *F. nucleatum*, and *T. forsythia* after treatment were not significantly different between probiotics treatment and control treatment groups at four-, and eight-week. Further forest plots representing the SMD of *P. gingivalis*, *P. intermedia*, *F. nucleatum*, and *T. forsythia* after probiotics treatment can be found in the publication of Sang-Ngoen *et al* (108).

3.1.7. Certainty of evidence assessment

The level of certainty of evidence was done using the GRADE approach on the efficacy of orally-administered probiotics to reduce the quantity of harmful periodontal bacteria. The outcomes from different pathogenic periodontal bacteria and duration of the measurement revealed very low grades in all assessments due to the presence of serious a risk of bias, inconsistency and imprecision. The summary of GRADE approaches be found in the publication of Sang-Ngoen *et al* (108).

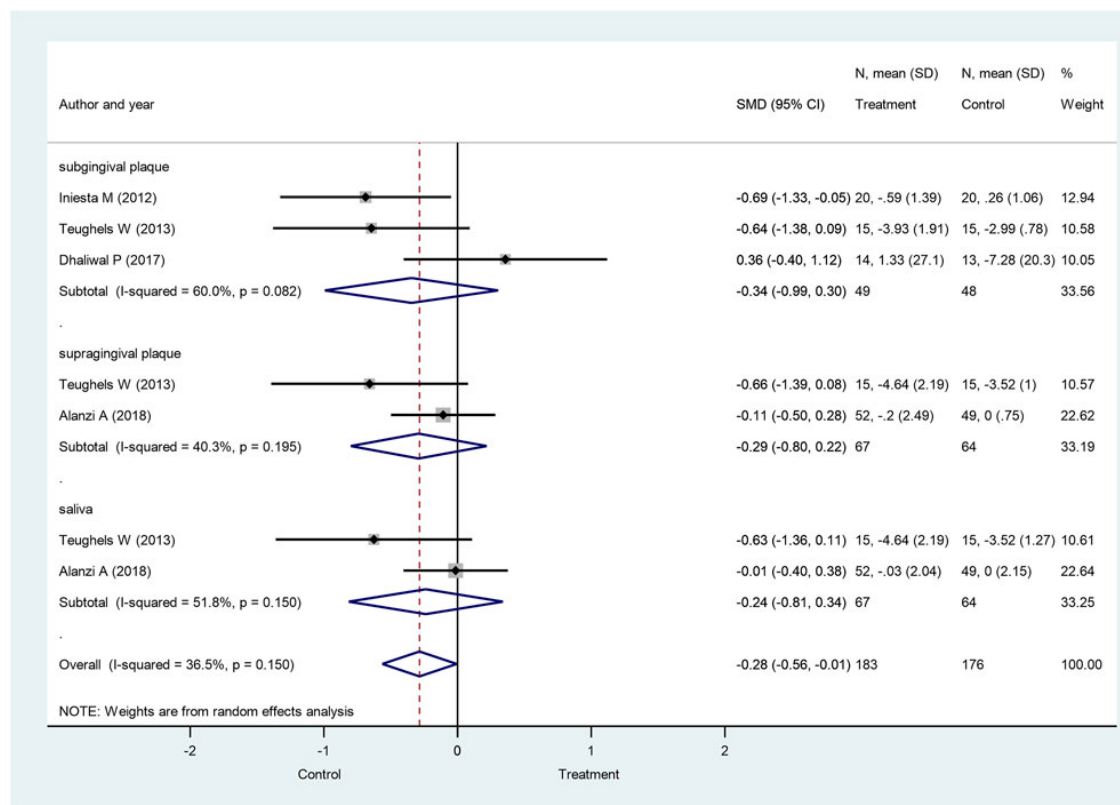


Figure 5. Forest plot analysis of the change in *A. actinomycetemcomitans* at 4 weeks (108).

The overall result of the standardized mean difference indicated a significant decrease of *A. actinomycetemcomitans* in the probiotic treatment over the control.

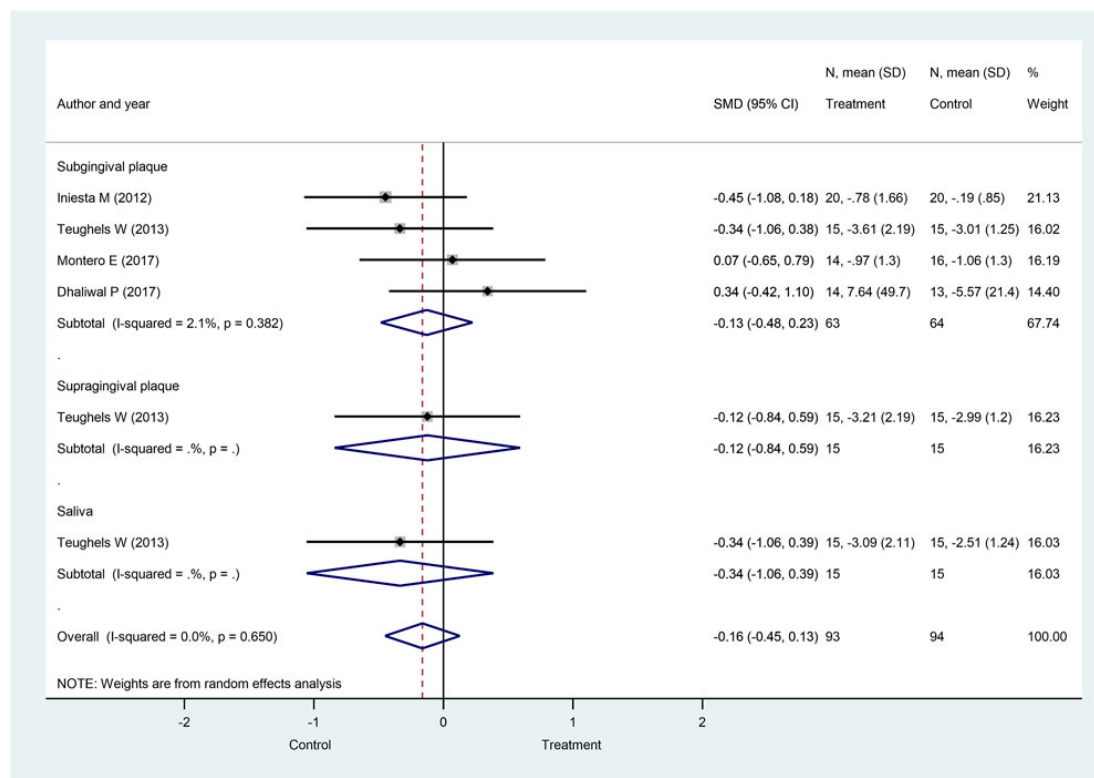


Figure 6. Forest plot analysis of the change in *A. actinomycetemcomitans* at 8 weeks (108).

The overall result of the standardized mean difference indicated a significant decrease of *A. actinomycetemcomitans* in the probiotic treatment over the control.

3.2. The efficacy of gabapentinoids to reduce acute herpes zoster pain occurrence

3.2.1. Protocol and registration

This meta-analysis protocol was registered in the PROSPERO; the registration number is CRD42018095758.

3.2.2. Clinical question

The PICO framework was constructed to investigate the efficacy of gabapentinoids on the reduction of the acute herpes zoster pain occurrence. The population (P) is patients with herpes zoster infection; the intervention (I) is gabapentinoids; the control (C) is placebo treatment or no treatment; and the outcome (O) is the presence of acute herpes zoster pain. The keywords used for the search was [*herpes zoster AND (gamma-aminobutyric acid OR gaba OR gabapentin OR neurontin OR pregabalin)*].

3.2.3. *The result of literature search and the selection of study*

The comprehensive literature search from PubMed, Web of Science, Ovid, Scopus and EMBASE databases yielded 4,888 records. No additional record was found through the bibliographic references of review articles. After removing duplicates, there were 3,130 records remaining and they were screened for titles and abstracts. Six records were relevant to the meta-analysis. Out of these, three records were excluded because one was the uncontrolled trial; another was a retrospective study (109, 110); while the third was an ongoing trial (111). The remaining three suitable records were extracted for data (112-114). See Figure 6 (below).

3.2.4. *Characteristics of included studies*

All included studies were single center RCTs (112-114). Each of studies exhibited some differences among each other such as the age of participants, the dosage and duration of treatment, and the use of antiviral agent. Only one study reported additional outcome on the quality of life (112). The summary characteristics of the included articles are presented in the publication of Sadaeng *et al* (115).

3.2.5. *The risk of bias assessment*

Although all studies stated that they were randomized trials, none of them detailed the method of randomization (112-114). Therefore, the selection bias in both random sequence generation and the allocation concealment were rated as a questionable risk of bias. All studies provided insufficient information on the blinding of participants and personnel, thus, they were determined as a questionable risk of bias in the domain of performance bias. A description of blinding outcome assessment method was described only in one study (112), therefore, it exhibited a low risk of bias in the domain of detection bias. Two studies were judged to have a high rate of incomplete outcome data (attrition bias domain) because of a high drop out rate and no dropout reason (112, 113). A low risk of reporting bias was determined in two studies (112, 113), while another was rated as a high risk because all pre-specified outcomes were not reported (114). To sum up, all studies exhibited a high risk of bias because there were at least one high risk of bias in the key domains. The results of risk of bias assessment are summarized and detailed in Figure 7.

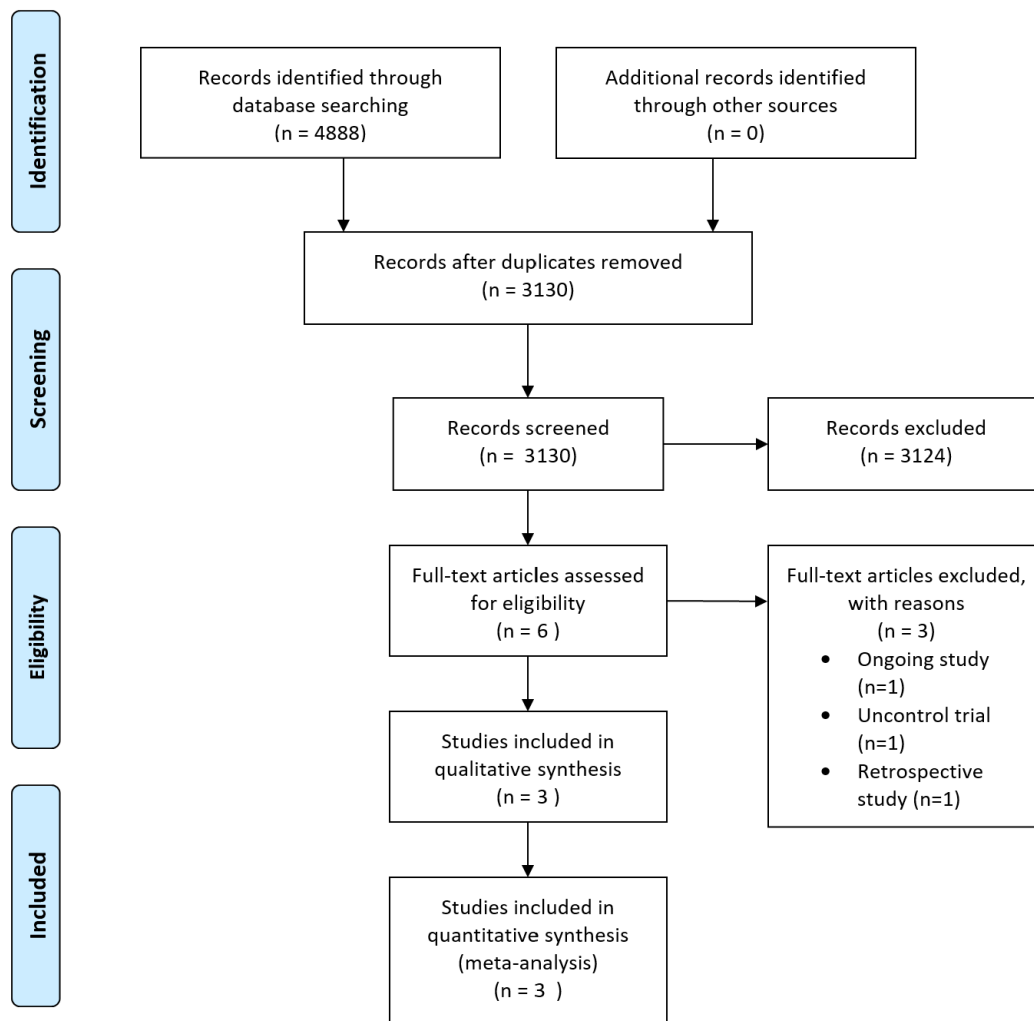


Figure 7. PRISMA 2009 flow diagram for identification of relevant studies (115).

	Random sequence generation (selection bias)	Allocation concealment (selection bias)	Blinding of participants and personnel (performance bias)	Blinding of outcome assessment (detection bias)	Incomplete outcome data (attrition bias)	Selective reporting (reporting bias)	Other bias
Lee E.G. et al. 2016	?	?	?	+	-	+	+
Skvarc N. et al 2010	?	?	?	?	-	+	+
Wang Q et al 2013	?	?	?	?	?	-	?

Figure 8. A summary of the risk of bias of included studies (115).

3.2.6. The result of meta-analysis

The presence of acute zoster pain after treatment

The forest plot showed the pooled odds ratios of events occurred after gabapentinoids treatment (Figure 9). The result indicated that the occurrence of acute herpes zoster pain in the gabapentinoids group was significantly lower compared to the placebo group (OR=0.36; 95% CI= 0.14 to 0.93; $p=0.035$; heterogeneity: $I^2=40.7\%$, $p=0.186$). This suggested that gabapentinoids could prevent acute zoster pain in patients after herpes zoster infection.

Presences of adverse events during treatment

The adverse events were noted in only two studies (112, 113). They reported different aspects of adverse events, but they shared some common aspect such as fatigue, constipation, and dizziness. There was no difference between the gabapentinoids group and the control group. The detail of adverse events was shown in the publication of Sadaeng *et al* (115). The meta-analysis was not performed due to insufficient data.

The quality of life

The dermatologic life quality index was assessed and reported in only one study (112). Patients suffering from HZI were asked to determine how their skin problem affects their quality of life. The quality of life between the gabapentinoids treated group and the control group was not significantly different. The meta-analysis on this basis was not performed due to insufficient data.

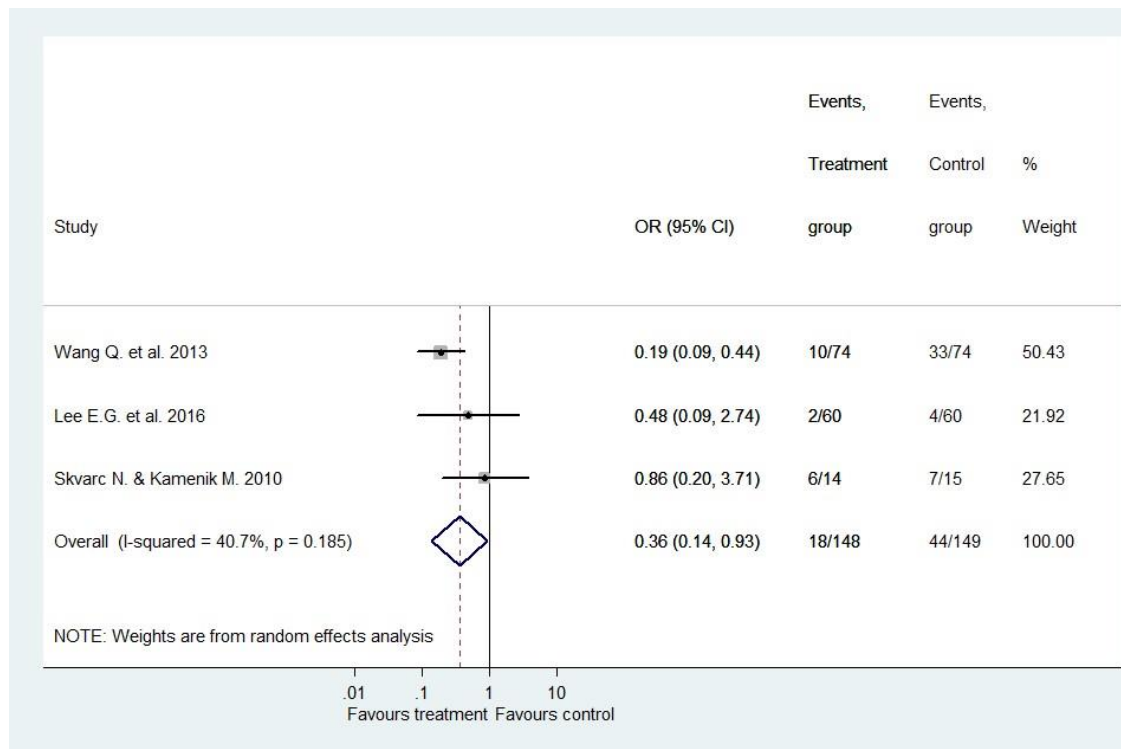


Figure 9. Forest plot analysis of the presence of acute zoster pain (115).

Overall results of odds ratio indicated a preventive effect from acute zoster pain in the gabapentinoids group over the placebo group.

3.2.7. Certainty of evidence assessment

The level of certainty of evidence was done using the GRADE approach on the efficacy of gabapentinoids to reduce the acute herpes zoster pain occurrence. The result reveled a very low grade because of the presence of a very high risk of bias and publication bias. The summary of GRADE approach is shown in the publication of Sadaeng *et al* (115).

4. Discussions

4.1. The efficacy of orally-administered probiotics to reduce the quantity of pathogenic periodontal bacteria

This meta-analysis aimed to evaluate whether orally administered probiotics decreases the level of pathogenic periodontal bacteria in patients diagnosed with periodontal diseases. The present study focused on five pathogenic bacteria which are highly related to periodontal disease progression and severity. The results from this meta-analysis suggested that the probiotics-treated group had a significant decrease of *A. actinomycetemcomitans* count when compared to the control groups at four weeks, but not at 8 weeks, after treatment initiation. Likewise, bacterial counts of *P. gingivalis*, *P. intermedia*, *F. nucleatum*, and *T. forsythia* at four weeks and eight weeks after treatment were not significantly different between probiotics and control groups.

Two studies, which included only two trials each, showed similar results to the results obtained in this study, suggesting that the use of probiotics has an insignificant effect in the reduction in the number of pathogenic periodontal bacteria count. In addition, a decreasing trend in the number of *A. actinomycetemcomitans*, *P. gingivalis*, *P. intermedia*, *F. nucleatum* and *T. forsythia* has been noted (116, 117). Other than the alteration of bacterial count, the clinical parameters after the probiotic used were also investigated and the results were inconsistent and diverse in each clinical parameter (70, 71, 88, 116-118). Diverse outcomes of clinical periodontal parameters have also been reported in reviews, either supporting or questioning the effectiveness of probiotics in periodontal diseases (65, 69, 72, 119-121). These controversial results may be explained by different follow-up times, various probiotic strains, and the dosage of probiotics in each trial, and different patients' characteristics.

The spectrum of periodontal diseases ranges from locally mild inflammation to generalized severe inflammation of the periodontium. In this study, all types of periodontal diseases were included for analyses. Also, all strains of probiotics were eligible for the study in order to yield the conclusive answer of the study question. Among the included studies, *Lactobacillus reuteri* has been frequently used as probiotics because it has a potency to overcome the pathogenic microorganisms by producing antimicrobial substances such as reuterin, reutericyclin, and lactic acid, and modulating the immune system (122). Additionally, *in vivo* experiments have supported the antimicrobial effect

of *L. reuteri* against cariogenic and periodontogenic bacteria (123-125). In harmony with these antimicrobial activities, included studies using *L. reuterin* found a reduction of periodontal pathogens, such as *A. actinomycetemcomitans*, *P. gingivalis*, and *T. forsythia*, in patients with periodontal diseases (63, 97, 100, 105). In contrast, the administration of *L. reuteri* had no effect over the periodontal pathogen at 12 and 24 weeks (102). This outcome could be reversed by using a mixture of *L. reuteri* with *Lactobacillus rhamnosus*, *Bifidobacterium longum*, and *Bifidobacterium bifidum* (97). Therefore, it might suggest that a mixture of *L. reuteri* with other beneficial bacteria could overcome the limitation of antibacterial effect in a long-term used of *L. reuteri* alone.

The efficacy of probiotics probably relied on many factors, including bacterial strains, dosages of probiotics, durations of treatment and follow-up time (126-128). Different probiotic species may affect differently on pathogenic bacteria (128, 129), and a specific pathogenic bacteria may probably be susceptible to a specific species of probiotics (128). In periodontitis, different pathogenic bacterial species has been identified to cause different types of periodontal diseases, ranging from mild to severe and acute to chronic inflammation (130, 131). Therefore, the selection of probiotics to match the type of periodontal diseases could be considered for future studies.

Besides the species of probiotics, the amount of probiotics used can influence the outcome. The acceptable dose of probiotics used per day is at the total amount of 10^8 - 10^9 microorganisms (126). Included studies administered the dose of probiotics within this range.

This present meta-analysis has unavoidable limitations. A low number of included articles with relative heterogeneity is the major limitation. A major heterogeneity of included studies is microbiological assessment between conventional cultivation and molecular PCR. Since a cultivation of periodontal pathogenic bacteria is difficult to perform and has low sensitivity (132-135). This could result in a lower number of bacterial counts compared to the PCR method. Moreover, all spectrum of periodontal diseases ranging from gingivitis to severe chronic periodontitis were included in this study. Other limitations were different probiotic strains, doses and forms, specific interaction between probiotic strains and pathogenic bacterial species, different genetic background and different environment such as oral hygiene procedure or pre-treatment

with scaling and root planning. These varieties across studies may have influenced the results.

4.2. The efficacy of gabapentinoids to reduce the acute herpes zoster pain occurrence

This meta-analysis aimed to investigate the effectiveness of gabapentinoids in reducing acute pain occurrence in patients who have HZV infection. By comparing the number of patients with the presence of pain between gabapentinoid-treated and placebo-treated groups, the pooled odds ratio indicated that the treatment of gabapentinoids significantly reduced the number of patients with acute zoster pain.

The pathophysiology of pain-associated with HZV infection involves both central and peripheral nervous system, resulting from the destruction of nervous system during the acute infection (136, 137). At the cellular level, the TRPV1 is upregulated and it is associated with pain (74). Furthermore, the expression of voltage-gated sodium channels and voltage-gated potassium channels is increased (138). As a consequence, peripheral nerves lose their ability to suppress nociceptive pain signals, thereby, decreasing the threshold of nociceptive sensory activation and producing spontaneous ectopic discharges. Then, peripheral and central sensitizations evoke, causing allodynia and hyperalgesia sensation (139). These pathologic outcomes cause neuropathic pain which can be controlled by GABA-like substances called gabapentinoids. A study of VZV induced neuropathic changes can be attenuated by the gabapentin or by sodium channel blockers (138).

Few well-designed studies investigating gabapentinoids were found in our preliminary literature search. Therefore, our PICO was planned to pool all types of gabapentinoids and compare them to the placebo by ignoring their differences. However, different derivatives diversely characterize in potency, adverse events, pharmacokinetics and pharmacodynamics (80). Two of included studies administered gabapentin (112, 114), while the remaining used pregabalin (113). The dosage used in included studies for gabapentin was 900 mg/day, and for pregabalin was 150 mg/day. All dosage included in this study were in a range of other trials using for PHN: 900 – 3,600 mg/day for gabapentin (84, 140-147), 150 – 600 mg/day for pregabalin (81, 148-155).

Two meta-analyses on adverse events of gabapentin and pregabalin indicated that the risk of adverse reactions significantly increased when the dosage increased (156, 157).

The common complaint of adverse events are somnolence, dizziness, ataxia, fatigue and peripheral edema which persuade patients to withdraw from the treatment (158, 159). These events usually occurred at the beginning of the treatment and at higher dosages. Therefore, a slow dose escalation was suggested to reduce the occurrence of adverse events. Included studies showed no serious adverse events of gabapentinoids. Only the study of Krčevski Škvarč *et al.* noted a significant difference in dizziness and somnolence between the pregabalin treated group and placebo groups (113).

The ability to control pain during herpes zoster infection may reduce the magnitude of the initiation phase of nociceptor evoked peripheral and central sensitivity. It was recommended to use an antiviral with effective pain control to better decrease the risk of persistent pain development than administering antiviral drugs alone (45). A published RCT protocol suggests starting the administration of gabapentin at the same time as starting antiviral therapy (160). However, the study of Bulilete *et al.* indicated that the antiviral agent with the additional gabapentin administration within 72 hours of rash onsets provided no significant relief from acute herpes zoster pain or prevent the further chronic pain (161). In the included trials, the administration of gabapentin or pregabalin was not started at the same time as the onset of the disease. We have to note that the initiation time of treatment might affect the results of individual studies and the following meta-analyses.

There are significant limitations to the present study. This study included a low number of studies with a high risk of bias, and a low number of participants. The studies have no clear randomization method and allocation concealment. This may result in imbalanced known and unknown risk factors and covariates in test and control group and selection bias. Heterogeneity of the included studies is high due to different methodologies between each study. For example, one study included participants older than 50 years old (112), while another study included participants age range from 30 – 80 years old (113). Furthermore, participants from one study received antiviral agents with the study drug within 72 hour after the onset of zoster rash (112), whereas participants in another study received treatment in a delay period (7-14 days after the onset of rash) (113). Accordingly, the results should be interpreted considering described limitations.

5. Conclusion

5.1. Meta-analysis of the efficacy of orally-administered probiotics to reduce the quantity of pathogenic periodontal bacteria

The results from our meta-analysis suggested that orally administered probiotics have an ability to reduce numbers of pathogenic periodontal bacteria – *A. actinomycetemcomitans* at 4 week, but not at 8 week, after the initiation of treatment in a pooled analysis. The amount of *A. actinomycetemcomitans* count in subgroup analyses from sub-gingiva, supra-gingiva and saliva tends to decrease after probiotics treatment. However, probiotics have no beneficial effect in reducing *P. gingivalis*, *P. intermedia*, *F. nucleatum*, and *T. forsythia*. The use of orally administered probiotics as an adjunct to the conventional scaling and root planning could reduce a specific strain of periodontal pathogenic bacteria in healthy periodontal patients. However, due to distinct heterogeneity among the available RCTs, standardized clinical protocol is needed to further evaluate the effect of various probiotics on periodontal pathogens.

5.2. Meta-analysis of the efficacy of gabapentinoids to reduce the acute herpes zoster pain occurrence

The results from our meta-analysis indicated that administration of gabapentinoids reduce the occurrence of acute herpes zoster pain after the healing of rash. Gabapentinoids also help improving the quality of life in patients who have suffered from pain during and after herpes zoster infection. However, patients could experience some adverse events, such as dizziness, which may be reasons for refusing treatment. This study provides preliminary evidence in the prevention of the development of PHN from the anti-sensitization aspect as the use of gabapentinoids can control pain which leads to the prevention of pain sensation. However, currently available evidence on this matter is weak. Therefore, additional, well designed randomized clinical trials are needed, as well as a long-term study with a higher dosage of gabapentinoids.

6. Summary

Pain is an unpleasant sensation and emotional experience of individuals. Generally, pain is perceived after tissue damage and infection, which stimulate immune cells to secrete cytokines. As a consequence, the inflammation is initiated and pain signal is generated from activated and sensitized nociceptors. Furthermore, nociceptive neurons can be activated and sensitized directly from microbes.

P. gingivalis, an anaerobic Gram-negative bacterium typically found in the oral cavity, is a causative agent for periodontitis. Its lipopolysaccharide can stimulate and sensitize nociceptors, resulting in pain. Thus, the first aim of our study was to investigate the ability of probiotics to reduce the presence of periodontal pathogenic microbial in periodontal diseases in order to decrease infection and pain levels.

Varicella-zoster virus (VZV), one of the herpes viruses found in the oral cavity, causes severe pain due to nerve damage, structural and functional alterations in the peripheral and central nervous system. Neuropathic pain caused by VZV infection can be controlled by the use of gabapentinoids. These agents block calcium channels which probably intercept the neuronal sensitization. Therefore, the second aim was to investigate the effect of gabapentinoids in reducing the occurrence of herpes zoster pain after the condition has healed.

To perform these studies, meta-analysis was used to evaluate the available evidence. Databases were searched for eligible studies. Then, the data were collected and analyzed to address each aim.

The results from the first meta-analysis showed that probiotics can significantly reduce the number of *A. actinomycetemcomitans* in patients with periodontal disease at 4 weeks, while other periodontal pathogens – *P. gingivalis*, *P. intermedia*, *F. nucleatum*, and *T. forsythia* – were not changed after the use of probiotics. The different response of pathogens and probiotics may probably arise from the specificity of probiotics-pathogens. The second meta-analysis on gabapentinoids and herpes zoster pain indicated that gabapentinoids can reduce the observed rate of recurring herpes zoster pain after the healing of acute VZV infection. However, limitations from inhomogeneity of study design, low number of included studies and participants, publication bias, and small study effects may influence the outcomes of meta-analyses. Therefore, additional, well-designed randomized clinical trials with large number of participants are needed to overcome those limitations.

In conclusion, periodontal disease-associated pain may be controlled by using specific probiotic. While, herpes zoster-associated pain can be managed with gabapentinoids, which could be beneficial in preventing the development of postherpetic neuralgia.

7. Bibliography

1. Raja SN, Carr DB, Cohen M, Finnerup NB, Flor H, Gibson S, Keefe FJ, Mogil JS, Ringkamp M, Sluka KA, Song X-J, Stevens B, Sullivan MD, Tutelman PR, Ushida T, Vader K. (2020) The revised International Association for the Study of Pain definition of pain: concepts, challenges, and compromises. *Pain*, 161: 1976-1982.
2. Kidd BL, Urban LA. (2001) Mechanisms of inflammatory pain. *Br. J. Anaesth.*, 87: 3-11.
3. Devor M. Neuropathic pain: Pathophysiological response of nerves to injury. In: McMahon SB, Koltzenburg M, Tracey I, Turk D (szerk.), Wall and Melzack's Textbook of Pain. Elsevier Churchill Livingstone, Philadelphia, 2013: 861-888.
4. Scholz J, Woolf CJ. (2002) Can we conquer pain? *Nat. Neurosci.*, 5: 1062-1067.
5. Tosi MF. (2005) Innate immune responses to infection. *J. Allergy Clin. Immunol.*, 116: 241-249.
6. Galicia JC, Henson BR, Parker JS, Khan AA. (2016) Gene expression profile of pulpitis. *Genes Immun.*, 17: 239-243.
7. Turner MD, Nedjai B, Hurst T, Pennington DJ. (2014) Cytokines and chemokines: At the crossroads of cell signalling and inflammatory disease. *Biochim. Biophys. Acta*, 1843: 2563-2582.
8. Werner S, Grose R. (2003) Regulation of Wound Healing by Growth Factors and Cytokines. *Physiol. Rev.*, 83: 835-870.
9. Lindell DM, Lukacs NW. Cytokines and Chemokines in Inflammation. In: Serhan CN, Gilroy DW, Ward PA (szerk.), *Fundamentals of Inflammation*. Cambridge University Press, Cambridge, 2010: 175-185.
10. Opal SM, DePalo VA. (2000) Anti-Inflammatory Cytokines. *Chest*, 117: 1162-1172.
11. Zhang J-M, An J. (2007) Cytokines, Inflammation, and Pain. *Int. Anesthesiol. Clin.*, 45: 27-37.
12. Ji RR, Chamech A, Zhang YQ. (2016) Pain regulation by non-neuronal cells and inflammation. *Science*, 354: 572-577.
13. Pinho-Ribeiro FA, Verri WA, Jr., Chiu IM. (2017) Nociceptor Sensory Neuron-Immune Interactions in Pain and Inflammation. *Trends Immunol.*, 38: 5-19.

14. Binshtok AM, Wang H, Zimmermann K, Amaya F, Vardeh D, Shi L, Brenner GJ, Ji RR, Bean BP, Woolf CJ, Samad TA. (2008) Nociceptors are interleukin-1beta sensors. *J. Neurosci.*, 28: 14062-14073.
15. Ji RR, Samad TA, Jin SX, Schmoll R, Woolf CJ. (2002) p38 MAPK activation by NGF in primary sensory neurons after inflammation increases TRPV1 levels and maintains heat hyperalgesia. *Neuron*, 36: 57-68.
16. Zhuang ZY, Gerner P, Woolf CJ, Ji RR. (2005) ERK is sequentially activated in neurons, microglia, and astrocytes by spinal nerve ligation and contributes to mechanical allodynia in this neuropathic pain model. *Pain*, 114: 149-159.
17. Caviedes-Bucheli J, Muñoz HR, Azuero-Holguín MM, Ulate E. (2008) Neuropeptides in Dental Pulp: The Silent Protagonists. *J. Endod.*, 34: 773-788.
18. Basbaum AI, Bautista DM, Scherrer G, Julius D. (2009) Cellular and Molecular Mechanisms of Pain. *Cell*, 139: 267-284.
19. Chiu IM. (2018) Infection, Pain, and Itch. *Neurosci. Bull.*, 34: 109-119.
20. Deng L, Chiu IM. (2021) Microbes and pain. *PLoS Pathog.*, 17: e1009398: 1-7.
21. Netea MG, Balkwill F, Chonchol M, Cominelli F, Donath MY, Giamarellos-Bourboulis EJ, Golenbock D, Gresnigt MS, Heneka MT, Hoffman HM, Hotchkiss R, Joosten LAB, Kastner DL, Korte M, Latz E, Libby P, Mandrup-Poulsen T, Mantovani A, Mills KHG, Nowak KL, O'Neill LA, Pickkers P, van der Poll T, Ridker PM, Schalkwijk J, Schwartz DA, Siegmund B, Steer CJ, Tilg H, van der Meer JWM, van de Veerdonk FL, Dinarello CA. (2017) A guiding map for inflammation. *Nat. Immunol.*, 18: 826-831.
22. Deo PN, Deshmukh R. (2019) Oral microbiome: Unveiling the fundamentals. *J. Oral Maxillofac. Pathol.*, 23: 122-128.
23. Canut-Delgado N, Giovannoni ML, Chimenos-Küstner E. (2021) Are probiotics a possible treatment of periodontitis? Probiotics against periodontal disease: a systematic review. *Br. Dent. J.*: 1-7.
24. Kinane DF, Stathopoulou PG, Papapanou PN. (2017) Periodontal diseases. *Nat Rev Dis Primers*, 3: 17038: 1-14.
25. Costalonga M, Herzberg MC. (2014) The oral microbiome and the immunobiology of periodontal disease and caries. *Immunol. Lett.*, 162: 22-38.

26. Darveau RP. (2010) Periodontitis: a polymicrobial disruption of host homeostasis. *Nat. Rev. Microbiol.*, 8: 481-490.
27. Hajishengallis G. (2014) Immunomicrobial pathogenesis of periodontitis: keystones, pathobionts, and host response. *Trends Immunol.*, 35: 3-11.
28. Roberts FA, Darveau RP. (2002) Beneficial bacteria of the periodontium. *Periodontol.* 2000, 30: 40-50.
29. Gendron R, Grenier D, Maheu-Robert L-F. (2000) The oral cavity as a reservoir of bacterial pathogens for focal infections. *Microbes Infect*, 2: 897-906.
30. Peres MA, Macpherson LMD, Weyant RJ, Daly B, Venturelli R, Mathur MR, Listl S, Celeste RK, Guarnizo-Herreño CC, Kearns C, Benzian H, Allison P, Watt RG. (2019) Oral diseases: a global public health challenge. *Lancet*, 394: 249-260.
31. Nazir MA. (2017) Prevalence of periodontal disease, its association with systemic diseases and prevention. *Int J Health Sci (Qassim)*, 11: 72-80.
32. Miranda-Rius J, Brunet-Llobet L, Lahor-Soler E. (2018) The Periodontium as a Potential Cause of Orofacial Pain: A Comprehensive Review. *Open Dent J*, 12: 520-528.
33. Brunsvold MA, Nair P, Oates TW. (1999) Chief complaints of patients seeking treatment for periodontitis. *J. Am. Dent. Assoc.*, 130: 359-364.
34. Lagomarsino VN, Kostic AD, Chiu IM. (2021) Mechanisms of microbial–neuronal interactions in pain and nociception. *Neurobiol Pain*, 9: 100056.
35. Wadachi R, Hargreaves KM. (2006) Trigeminal nociceptors express TLR-4 and CD14: a mechanism for pain due to infection. *J. Dent. Res.*, 85: 49-53.
36. Diogenes A, Ferraz CC, Akopian AN, Henry MA, Hargreaves KM. (2011) LPS sensitizes TRPV1 via activation of TLR4 in trigeminal sensory neurons. *J. Dent. Res.*, 90: 759-764.
37. Meseguer V, Alpizar YA, Luis E, Tajada S, Denlinger B, Fajardo O, Manenschijn JA, Fernandez-Pena C, Talavera A, Kichko T, Navia B, Sanchez A, Senaris R, Reeh P, Perez-Garcia MT, Lopez-Lopez JR, Voets T, Belmonte C, Talavera K, Viana F. (2014) TRPA1 channels mediate acute neurogenic inflammation and pain produced by bacterial endotoxins. *Nat Commun*, 5: 3125: 1-14.

38. Ferraz CCR, Henry MA, Hargreaves KM, Diogenes A. (2011) Lipopolysaccharide From *Porphyromonas gingivalis* Sensitizes Capsaicin-Sensitive Nociceptors. J. Endod., 37: 45-48.
39. Asai D, Nakashima H. (2018) Pathogenic Viruses Commonly Present in the Oral Cavity and Relevant Antiviral Compounds Derived from Natural Products. Medicines, 5: 120: 1-18.
40. Slots J. (2015) Periodontal herpesviruses: prevalence, pathogenicity, systemic risk. Periodontol. 2000, 69: 28-45.
41. Gershon AA, Breuer J, Cohen JI, Cohrs RJ, Gershon MD, Gilden D, Grose C, Hambleton S, Kennedy PGE, Oxman MN, Seward JF, Yamanishi K. (2015) Varicella zoster virus infection. Nat Rev Dis Primers, 1: 15016: 1-18.
42. Kennedy PGE, Gershon AA. (2018) Clinical Features of Varicella-Zoster Virus Infection. Viruses, 10: 609: 1-11.
43. Warwick RA, Hanani M. (2016) Involvement of aberrant calcium signalling in herpetic neuralgia. Exp. Neurol., 277: 10-18.
44. Duncan CJA, Hambleton S. (2015) Varicella zoster virus immunity: A primer. J. Infect., 71: S47-S53.
45. Dworkin RH, Schmader KE, Goldstein EJC. (2003) Treatment and Prevention of Postherpetic Neuralgia. Clin. Infect. Dis., 36: 877-882.
46. Hadley GR, Gayle JA, Ripoll J, Jones MR, Argoff CE, Kaye RJ, Kaye AD. (2016) Post-herpetic Neuralgia: a Review. Curr Pain Headache Rep, 20: 17: 1-5.
47. Schmader KE, Dworkin RH. The Epidemiology and Natural History of Herpes Zoster and Postherpetic Neuralgia. In: Watson CPN, Gershon AA, Oxman MN (szerk.), Herpes Zoster: Postherpetic Neuralgia and Other Complications: Focus on Treatment and Prevention. Springer International Publishing, Cham, 2017: 25-44.
48. Gan EY, Tian EAL, Tey HL. (2013) Management of Herpes Zoster and Post-Herpetic Neuralgia. Am. J. Clin. Dermatol., 14: 77-85.
49. Mallick-Searle T, Snodgrass B, Brant JM. (2016) Postherpetic neuralgia: epidemiology, pathophysiology, and pain management pharmacology. J Multidiscip Healthc, 9: 447-454.

50. Dworkin RH, Portenoy RK. (1996) Pain and its persistence in herpes zoster. *Pain*, 67: 241-251.
51. Kress M, Fickenscher H. (2001) Infection by human varicella-zoster virus confers norepinephrine sensitivity to sensory neurons from rat dorsal root ganglia. *FASEB J.*, 15: 1037-1043.
52. Park CK, Xu ZZ, Berta T, Han Q, Chen G, Liu XJ, Ji RR. (2014) Extracellular microRNAs activate nociceptor neurons to elicit pain via TLR7 and TRPA1. *Neuron*, 82: 47-54.
53. Mayer M, James M, Russell R, Kelly J, Pasternak C. (1986) Changes in excitability induced by herpes simplex viruses in rat dorsal root ganglion neurons. *J. Neurosci.*, 6: 391-402.
54. Zappa U, Smith B, Simona C, Graf H, Case D, Kim W. (1991) Root Substance Removal by Scaling and Root Planing. *J. Periodontol.*, 62: 750-754.
55. Claffey N, Polyzois I, Ziaka P. (2004) An overview of nonsurgical and surgical therapy. *Periodontol.* 2000, 36: 35-44.
56. Cobb CM. (2002) Clinical significance of non-surgical periodontal therapy: an evidence-based perspective of scaling and root planing. *J. Clin. Periodontol.*, 29: 22-32.
57. Cobb CM. (2008) Microbes, inflammation, scaling and root planing, and the periodontal condition. *J. Dent. Hyg.*, 82 Suppl 3: 4-9.
58. McGowan K, McGowan T, Ivanovski S. (2018) Optimal dose and duration of amoxicillin-plus-metronidazole as an adjunct to non-surgical periodontal therapy: A systematic review and meta-analysis of randomized, placebo-controlled trials. *J. Clin. Periodontol.*, 45: 56-67.
59. Allaker RP, Stephen AS. (2017) Use of Probiotics and Oral Health. *Current Oral Health Reports*, 4: 309-318.
60. Devine DA, Marsh PD. (2009) Prospects for the development of probiotics and prebiotics for oral applications. *J. Oral Microbiol.*, 1: 1949: 1-11.
61. Meimandi M, Talebi Ardakani MR, Esmaeilnejad A, Yousefnejad P, Saebi K, Tayeed MH. (2017) The Effect of Photodynamic Therapy in the Treatment of Chronic Periodontitis: A Review of Literature. *Journal of Lasers in Medical Sciences*, 8: S7-S11.

62. Sanders ME. (2008) Probiotics: Definition, Sources, Selection, and Uses. Clin. Infect. Dis., 46: S58-S61.
63. Teughels W, Durukan A, Ozcelik O, Pauwels M, Quirynen M, Haytac MC. (2013) Clinical and microbiological effects of *Lactobacillus reuteri* probiotics in the treatment of chronic periodontitis: a randomized placebo-controlled study. J. Clin. Periodontol., 40: 1025-1035.
64. Tekce M, Ince G, Gursoy H, Dirikan Ipci S, Cakar G, Kadir T, Yılmaz S. (2015) Clinical and microbiological effects of probiotic lozenges in the treatment of chronic periodontitis: a 1-year follow-up study. J. Clin. Periodontol., 42: 363-372.
65. Seminario-Amez M, Lopez-Lopez J, Estrugo-Devesa A, Ayuso-Montero R, Jane-Salas E. (2017) Probiotics and oral health: A systematic review. Med. Oral Patol. Oral Cir. Bucal., 22: e282-e288.
66. Ahola AJ, Yli-Knuuttila H, Suomalainen T, Poussa T, Ahlström A, Meurman JH, Korpela R. (2002) Short-term consumption of probiotic-containing cheese and its effect on dental caries risk factors. Arch. Oral Biol., 47: 799-804.
67. Montalto M, Vastola M, Marigo L, Covino M, Graziosetto R, Curigliano V, Santoro L, Cuoco L, Manna R, Gasbarrini G. (2004) Probiotic Treatment Increases Salivary Counts of Lactobacilli: A Double-Blind, Randomized, Controlled Study. Digestion, 69: 53-56.
68. Montero E, Iniesta M, Rodrigo M, Marín MJ, Figuero E, Herrera D, Sanz M. (2017) Clinical and microbiological effects of the adjunctive use of probiotics in the treatment of gingivitis: A randomized controlled clinical trial. J. Clin. Periodontol., 44: 708-716.
69. Yanine N, Araya I, Brignardello-Petersen R, Carrasco-Labra A, González A, Preciado A, Villanueva J, Sanz M, Martin C. (2013) Effects of probiotics in periodontal diseases: a systematic review. Clin. Oral Investig., 17: 1627-1634.
70. Martin-Cabezas R, Davideau J-L, Tenenbaum H, Huck O. (2016) Clinical efficacy of probiotics as an adjunctive therapy to non-surgical periodontal treatment of chronic periodontitis: a systematic review and meta-analysis. J. Clin. Periodontol., 43: 520-530.

71. Akram Z, Shafqat S, Aati S, Kujan O, Fawzy A. (2020) Clinical efficacy of probiotics in the treatment of gingivitis: A systematic review and meta-analysis. *Aust. Dent. J.*, 65: 12-20.
72. Vives-Soler A, Chimenos-Kustner E. (2020) Effect of probiotics as a complement to non-surgical periodontal therapy in chronic periodontitis: a systematic review. *Med. Oral Patol. Oral Cir. Bucal.*, 25: e161-e167.
73. Backonja M-M, Serra J. (2004) Pharmacologic Management Part 1: Better-Studied Neuropathic Pain Diseases. *Pain Med.*, 5: S28-S47.
74. Baron R, Binder A, Wasner G. (2010) Neuropathic pain: diagnosis, pathophysiological mechanisms, and treatment. *Lancet Neurol*, 9: 807-819.
75. Nicholson BD. (2003) Diagnosis and management of neuropathic pain: a balanced approach to treatment. *J. Am. Acad. Nurse Pract.*, 15: 3-9.
76. Finnerup NB, Attal N, Haroutounian S, McNicol E, Baron R, Dworkin RH, Gilron I, Haanpää M, Hansson P, Jensen TS, Kamerman PR, Lund K, Moore A, Raja SN, Rice ASC, Rowbotham M, Sena E, Siddall P, Smith BH, Wallace M. (2015) Pharmacotherapy for neuropathic pain in adults: a systematic review and meta-analysis. *Lancet Neurol*, 14: 162-173.
77. Johnson RW, Rice ASC. (2014) Postherpetic Neuralgia. *N. Engl. J. Med.*, 371: 1526-1533.
78. Thakur R, Philip AG. (2012) Chronic pain perspectives: Treating herpes zoster and postherpetic neuralgia: an evidence-based approach. *J. Fam. Pract.*, 61: S9-15.
79. Makharita MY, Amr YM, El-Bayoumy Y. (2015) Single Paravertebral Injection for Acute Thoracic Herpes Zoster: A Randomized Controlled Trial. *Pain Pract*, 15: 229-235.
80. Taylor CP. (2009) Mechanisms of analgesia by gabapentin and pregabalin – Calcium channel $\alpha 2\text{-}\delta$ [Cav $\alpha 2\text{-}\delta$] ligands. *Pain*, 142: 13-16.
81. Bockbrader HN, Burger P, Knapp L, Corrigan BW. (2011) Population pharmacokinetics of pregabalin in healthy subjects and patients with chronic pain or partial seizures. *Epilepsia*, 52: 248-257.
82. Scheinfeld N. (2003) The role of gabapentin in treating diseases with cutaneous manifestations and pain. *Int. J. Dermatol.*, 42: 491-495.

83. Rose MA, Kam PCA. (2002) Gabapentin: pharmacology and its use in pain management. *Anaesthesia*, 57: 451-462.
84. Beydoun A. (1999) Postherpetic Neuralgia: Role of Gabapentin and Other Treatment Modalities. *Epilepsia*, 40: s51-s56.
85. Lee GI, Neumeister MW. (2020) Pain: pathways and physiology. *Clin. Plast. Surg.*, 47: 173-180.
86. Zahradnik RT, Magnusson I, Walker C, McDonell E, Hillman CH, Hillman JD. (2009) Preliminary assessment of safety and effectiveness in humans of ProBiora3™, a probiotic mouthwash. *J. Appl. Microbiol.*, 107: 682-690.
87. Iwamoto T, Suzuki N, Tanabe K, Takeshita T, Hirofuji T. (2010) Effects of probiotic *Lactobacillus salivarius* WB21 on halitosis and oral health: an open-label pilot trial. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*, 110: 201-208.
88. Imran F, Das S, Padmanabhan S, Rao R, Suresh A, Bharath D. (2015) Evaluation of the efficacy of a probiotic drink containing *Lactobacillus casei* on the levels of periodontopathic bacteria in periodontitis: A clinico-microbiologic study. *Indian J. Dent. Res.*, 26: 462-468.
89. Kaklamanos EG, Nassar R, Kalfas S, Al Halabi M, Kowash M, Hannawi H, Hussein I, Salami A, Hassan A, Senok AC. (2019) A single-centre investigator-blinded randomised parallel group clinical trial to investigate the effect of probiotic strains *Streptococcus salivarius* M18 and *Lactobacillus acidophilus* on gingival health of paediatric patients undergoing treatment with fixed orthodontic appliances: study protocol. *BMJ Open*, 9: e030638: 1-6.
90. Elsadek MF, Ahmed BM, Alkhawtani DM, Zia Siddiqui A. (2020) A comparative clinical, microbiological and glycemic analysis of photodynamic therapy and *Lactobacillus reuteri* in the treatment of chronic periodontitis in type-2 diabetes mellitus patients. *Photodiagnosis Photodyn. Ther.*, 29: 101629: 1-5.
91. Hallström H, Lindgren S, Yucel-Lindberg T, Dahlén G, Renvert S, Twetman S. (2013) Effect of probiotic lozenges on inflammatory reactions and oral biofilm during experimental gingivitis. *Acta Odontol. Scand.*, 71: 828-833.

92. Becirovic A, Abdi-Dezfuli JF, Hansen MF, Lie SA, Vasstrand EN, Bolstad AI. (2018) The effects of a probiotic milk drink on bacterial composition in the supra- and subgingival biofilm: a pilot study. *Benef Microbes*, 9: 865-874.
93. Boyeena L, Koduganti R, Panthula V, Jammula S. (2019) Comparison of efficacy of probiotics versus tetracycline fibers as adjuvants to scaling and root planing. *J. Indian Soc. Periodontol.*, 23: 539-544.
94. Tobita K, Watanabe I, Tomokiyo M, Saito M. (2018) Effects of heat-treated *Lactobacillus crispatus* KT-11 strain consumption on improvement of oral cavity environment: a randomised double-blind clinical trial. *Benef Microbes*, 9: 585-592.
95. Meenakshi SS, Varghese S. (2018) Adjunctive effect of probiotic (*Lactobacillus casei* Shirota) to scaling and root planing in the management of chronic periodontitis. *Drug Invent Today*, 10: 1381-1386.
96. Mayanagi G, Kimura M, Nakaya S, Hirata H, Sakamoto M, Benno Y, Shimauchi H. (2009) Probiotic effects of orally administered *Lactobacillus salivarius* WB21-containing tablets on periodontopathic bacteria: a double-blinded, placebo-controlled, randomized clinical trial. *J. Clin. Periodontol.*, 36: 506-513.
97. Goyal N, Shamanna P, Varughese S, Abraham R, Antony B, Emmatty R, Paul P. (2019) Effects of amine fluoride and probiotic mouthwash on levels of *Porphyromonas gingivalis* in orthodontic patients: A randomized controlled trial. *J. Indian Soc. Periodontol.*, 23: 339-344.
98. Shah MP, Gujjari SK, Chandrasekhar VS. (2013) Evaluation of the Effect of Probiotic (Inersan[®]) Alone, Combination of Probiotic with Doxycycline and Doxycycline Alone on Aggressive Periodontitis; A Clinical and Microbiological Study. *J Clin Diag Res*, 7: 595-600.
99. Shah M, Gujjari S, Chandrasekhar V. (2017) Long-term effect of *Lactobacillus brevis* CD2 (Inersan[®]) and/or doxycycline in aggressive periodontitis. *J. Indian Soc. Periodontol.*, 21: 341-343.
100. Vivekananda MR, Vandana KL, Bhat KG. (2010) Effect of the probiotic *Lactobacilli reuteri* (Prodentis) in the management of periodontal disease: a preliminary randomized clinical trial. *J. Oral Microbiol.*, 2: 5344: 1-9.

101. Laleman I, Yilmaz E, Ozcelik O, Haytac C, Pauwels M, Herrero ER, Slomka V, Quirynen M, Alkaya B, Teughels W. (2015) The effect of a streptococci containing probiotic in periodontal therapy: a randomized controlled trial. *J. Clin. Periodontol.*, 42: 1032-1041.
102. Laleman I, Pauwels M, Quirynen M, Teughels W. (2020) A dual-strain *Lactobacilli reuteri* probiotic improves the treatment of residual pockets: A randomized controlled clinical trial. *J. Clin. Periodontol.*, 47: 43-53.
103. Alanzi A, Honkala S, Honkala E, Varghese A, Tolvanen M, Söderling E. (2018) Effect of *Lactobacillus rhamnosus* and *Bifidobacterium lactis* on gingival health, dental plaque, and periodontopathogens in adolescents: a randomised placebo-controlled clinical trial. *Benef Microbes*, 9: 593-602.
104. Invernici MM, Salvador SL, Silva PHF, Soares MSM, Casarin R, Palioto DB, Souza SLS, Taba Jr M, Novaes Jr AB, Furlaneto FAC, Messoria MR. (2018) Effects of *Bifidobacterium* probiotic on the treatment of chronic periodontitis: A randomized clinical trial. *J. Clin. Periodontol.*, 45: 1198-1210.
105. Iniesta M, Herrera D, Montero E, Zurbriggen M, Matos AR, Marín MJ, Sánchez-Beltrán MC, Llama-Palacio A, Sanz M. (2012) Probiotic effects of orally administered *Lactobacillus reuteri*-containing tablets on the subgingival and salivary microbiota in patients with gingivitis. A randomized clinical trial. *J. Clin. Periodontol.*, 39: 736-744.
106. Dhaliwal PK, Grover V, Malhotra R, Kapoor A. (2017) Clinical and Microbiological Investigation of the Effects of Probiotics Combined with Scaling and Root Planing in the Management of Chronic Periodontitis: A Randomized, Controlled Study. *J. Int. Acad. Periodontol.*, 19: 101-108.
107. Morales A, Gandolfo A, Bravo J, Carvajal P, Silva N, Godoy C, Garcia-Sesnich J, Hoare A, Diaz P, Gamonal J. (2018) Microbiological and clinical effects of probiotics and antibiotics on nonsurgical treatment of chronic periodontitis: a randomized placebo- controlled trial with 9-month follow-up. *J Appl Oral Sci*, 26: e20170075: 1-9.
108. Sang-Ngoen T, Czumbel LM, Sadaeng W, Miko A, Nemeth DI, Matrai P, Hegyi P, Toth B, Csutor D, Kiss I, Szabo A, Gerber G, Varga G, Keremi B. (2021) Orally Administered Probiotics Decrease *Aggregatibacter*

- actinomycetemcomitans but Not Other Periodontal Pathogenic Bacteria Counts in the Oral Cavity: A Systematic Review and Meta-Analysis. *Front. Pharmacol.*, 12: 682656: 1-14.
109. Lapolla W, DiGiorgio C, Haitz K, Magel G, Mendoza N, Grady J, Lu W, Tyring S. (2011) Incidence of Postherpetic Neuralgia After Combination Treatment With Gabapentin and Valacyclovir in Patients With Acute Herpes Zoster: Open-label Study. *Arch. Dermatol.*, 147: 901-907.
 110. Migita T, Maekawa T, Okada K, Kobayashi M, Egi A. (2013) Can early administration of pregabalin reduce the incidence of postherpetic neuralgia? *Eur. J. Anaesthesiol.*, 30: 207-207.
 111. Rullán M, Bulilete O, Leiva A, Soler A, Roca A, González-Bals MJ, Lorente P, Llobera J, Cladera M, Comas C, Mir MA, Cifre A, Lliteras B, Gestoso S, Jover A, Bestard F, Comas F, López L, Ortuño R, Peiro J, Cerdó M, Ramírez V, Gutierrez M, Argüelles R, Gutierrez MD, group PHN. (2017) Efficacy of gabapentin for prevention of postherpetic neuralgia: study protocol for a randomized controlled clinical trial. *Trials*, 18:24: 1-9.
 112. Lee EG, Lee HJ, Hyun DJ, Min K, Kim DH, Yoon MS. (2016) Efficacy of low dose gabapentin in acute herpes zoster for preventing postherpetic neuralgia: a prospective controlled study. *Dermatol. Ther.*, 29: 184-190.
 113. Krčevski Škvarč N, Kamenik M. (2010) Effects of pregabalin on acute herpetic pain and postherpetic neuralgia incidence. *Wien. Klin. Wochenschr.*, 122: 49-53.
 114. Wang Q, Cui W, Song T. (2013) The effectiveness of Gabapentin for the prevention of postherpetic neuralgia in patients with acute herpes zoster. *J. Pharmacol. Sci.*, 121: 194P.
 115. Sadaeng W, Marta K, Matrai P, Hegyi P, Toth B, Nemeth B, Czumbel LM, Sang-Ngoen T, Gyongyi Z, Varga G, Revesz P, Szanyi I, Karadi K, Gerber G. (2020) gamma-Aminobutyric Acid and Derivatives Reduce the Incidence of Acute Pain after Herpes Zoster - A Systematic Review and Meta-analysis. *Curr. Pharm. Des.*, 26: 3026-3038.
 116. Gruner D, Paris S, Schwendicke F. (2016) Probiotics for managing caries and periodontitis: Systematic review and meta-analysis. *J. Dent.*, 48: 16-25.

117. Ho SN, Acharya A, Sidharthan S, Li KY, Leung WK, McGrath C, Pelekos G. (2020) A Systematic Review and Meta-analysis of Clinical, Immunological, and Microbiological Shift in Periodontitis After Nonsurgical Periodontal Therapy With Adjunctive Use of Probiotics. *J Evid Based Dent Pract*, 20: 101397: 1-14.
118. Ikram S, Hassan N, Raffat MA, Mirza S, Akram Z. (2018) Systematic review and meta-analysis of double-blind, placebo-controlled, randomized clinical trials using probiotics in chronic periodontitis. *J. Investig. Clin. Dent.*, 9: e12338: 1-9.
119. Barboza EP, Arriaga PC, Luz DP, Montez C, Vianna KC. (2020) Systematic review of the effect of probiotics on experimental gingivitis in humans. *Braz Oral Res*, 34: e031: 1-9.
120. Deepa D, Mehta DS. (2009) Is the role of probiotics friendly in the treatment of periodontal diseases !! *J. Indian Soc. Periodontol.*, 13: 30-31.
121. Matsubara VH, Bandara HMHN, Ishikawa KH, Mayer MPA, Samaranayake LP. (2016) The role of probiotic bacteria in managing periodontal disease: a systematic review. *Expert Rev. Anti Infect. Ther.*, 14: 643-655.
122. Britton RA. Chapter 8 - *Lactobacillus reuteri*. In: Floch MH, Ringel Y, Allan Walker W (szerk.), *The Microbiota in Gastrointestinal Pathophysiology*. Academic Press, Boston, 2017: 89-97.
123. Baca-Castañón ML, De la Garza-Ramos MA, Alcázar-Pizaña AG, Grondin Y, Coronado-Mendoza A, Sánchez-Najera RI, Cárdenas-Estrada E, Medina-De la Garza CE, Escamilla-García E. (2015) Antimicrobial Effect of *Lactobacillus reuteri* on Cariogenic Bacteria *Streptococcus gordonii*, *Streptococcus mutans*, and Periodontal Diseases *Actinomyces naeslundii* and *Tannerella forsythia*. *Probiotics Antimicrob Proteins*, 7: 1-8.
124. Kang M-S, Oh J-S, Lee H-C, Lim H-S, Lee S-W, Yang K-H, Choi N-K, Kim S-M. (2011) Inhibitory effect of *Lactobacillus reuteri* on periodontopathic and cariogenic bacteria. *J Microbiol*, 49: 193-199.
125. Santos TA, Scorzoni L, Correia R, Junqueira JC, Anbinder AL. (2020) Interaction between *Lactobacillus reuteri* and periodontopathogenic bacteria using in vitro and in vivo (*G. Mellonella*) approaches. *Pathog Dis*, 78: 1-8.
126. Ciorba MA. (2012) A Gastroenterologist's Guide to Probiotics. *Clin. Gastroenterol. Hepatol.*, 10: 960-968.

127. Kechagia M, Basoulis D, Konstantopoulou S, Dimitriadi D, Gyftopoulou K, Skarmoutsou N, Fakiri EM. (2013) Health Benefits of Probiotics: A Review. *ISRN Nutr*, 2013: 48165: 1-7.
128. Verna EC, Lucak S. (2010) Use of probiotics in gastrointestinal disorders: what to recommend? *Therap. Adv. Gastroenterol.*, 3: 307-319.
129. Schrezenmeir J, de Vrese M. (2001) Probiotics, prebiotics, and synbiotics—approaching a definition. *Am. J. Clin. Nutr.*, 73: 361s-364s.
130. Ardila C, Alzate J, Guzmán I. (2012) Relationship between Gram negative enteric rods, *Aggregatibacter actinomycetemcomitans*, and clinical parameters in periodontal disease. *J. Indian Soc. Periodontol.*, 16: 65-69.
131. Farias BC, Souza PR, Ferreira B, Melo RS, Machado FB, Gusmao ES, Cimoës R. (2012) Occurrence of periodontal pathogens among patients with chronic periodontitis. *Braz. J. Microbiol.*, 43: 909-916.
132. Boutaga K, van Winkelhoff AJ, Vandenbroucke-Grauls CMJE, Savelkoul PHM. (2005) Periodontal pathogens: A quantitative comparison of anaerobic culture and real-time PCR. *FEMS Immunol. Med. Microbiol.*, 45: 191-199.
133. Boutaga K, Winkelhoff AJv, Vandenbroucke-Grauls CMJE, Savelkoul PHM. (2003) Comparison of Real-Time PCR and Culture for Detection of *Porphyromonas gingivalis* in Subgingival Plaque Samples. *J. Clin. Microbiol.*, 41: 4950-4954.
134. Jervøe-Storm P-M, Koltzsch M, Falk W, Dörfler A, Jepsen S. (2005) Comparison of culture and real-time PCR for detection and quantification of five putative periodontopathogenic bacteria in subgingival plaque samples. *J. Clin. Periodontol.*, 32: 778-783.
135. Kotsilkov K, Popova C, Boyanova L, Setchanova L, Mitov I. (2015) Comparison of culture method and real-time PCR for detection of putative periodontopathogenic bacteria in deep periodontal pockets. *Biotechnology & Biotechnological Equipment*, 29: 996-1002.
136. Head H, Campbell AW, Kennedy PGE. (1997) The pathology of Herpes Zoster and its bearing on sensory localisation. *Rev. Med. Virol.*, 7: 131-143.
137. Argoff CE, Katz N, Backonja M. (2004) Treatment of postherpetic neuralgia: a review of therapeutic options. *J. Pain Symptom Manage.*, 28: 396-411.

138. Garry EM, Delaney A, Anderson HA, Sirinathsinghji EC, Clapp RH, Martin WJ, Kinchington PR, Krah DL, Abbadie C, Fleetwood-Walker SM. (2005) Varicella zoster virus induces neuropathic changes in rat dorsal root ganglia and behavioral reflex sensitisation that is attenuated by gabapentin or sodium channel blocking drugs. *Pain*, 118: 97-111.
139. Woolf Clifford J, Max Mitchell B. (2001) Mechanism-based Pain Diagnosis: Issues for Analgesic Drug Development. *Anesthesiology*, 95: 241-249.
140. Backonja M, Glanzman RL. (2003) Gabapentin dosing for neuropathic pain: Evidence from randomized, placebo-controlled clinical trials. *Clin. Ther.*, 25: 81-104.
141. Baron R. (2004) Post-herpetic neuralgia case study: optimizing pain control. *Eur. J. Neurol.*, 11: 3-11.
142. Irving G, Jensen M, Cramer M, Wu J, Chiang Y-K, Tark M, Wallace M. (2009) Efficacy and Tolerability of Gastric-retentive Gabapentin for the Treatment of Postherpetic Neuralgia: Results of a Double-blind, Randomized, Placebo-controlled Clinical Trial. *Clin J Pain*, 25: 185-192.
143. Rauck RL, Irving GA, Wallace MS, Vanhove GF, Sweeney M. (2013) Once-Daily Gastroretentive Gabapentin for Postherpetic Neuralgia: Integrated Efficacy, Time to Onset of Pain Relief and Safety Analyses of Data From Two Phase 3, Multicenter, Randomized, Double-Blind, Placebo-Controlled Studies. *J. Pain Symptom Manage.*, 46: 219-228.
144. Rice ASC, Maton S, Group PNS. (2001) Gabapentin in postherpetic neuralgia: a randomised, double blind, placebo controlled study. *Pain*, 94: 215-224.
145. Rowbotham M, Harden N, Stacey B, Bernstein P, Magnus-Miller L, Group ftGPNS. (1998) Gabapentin for the Treatment of Postherpetic NeuralgiaA Randomized Controlled Trial. *JAMA*, 280: 1837-1842.
146. Wallace MS, Irving G, Cowles VE. (2010) Gabapentin Extended-Release Tablets for the Treatment of Patients with Postherpetic Neuralgia. *Clin. Drug Investig.*, 30: 765-776.
147. Werner RN, Nikkels AF, Marinovic B, Schafer M, Czarnecka-Operacz M, Agius AM, Bata-Csorgo Z, Breuer J, Girolomoni G, Gross GE, Langan S, Lapid-Gortzak R, Lesser TH, Pleyer U, Sellner J, Verjans GM, Wutzler P, Dressler C,

- Erdmann R, Rosumeck S, Nast A. (2017) European consensus-based (S2k) Guideline on the Management of Herpes Zoster - guided by the European Dermatology Forum (EDF) in cooperation with the European Academy of Dermatology and Venereology (EADV), Part 2: Treatment. *J. Eur. Acad. Dermatol. Venereol.*, 31: 20-29.
148. Cappuzzo KA. (2009) Treatment of postherpetic neuralgia: focus on pregabalin. *Clin. Interv. Aging*, 4: 17-23.
 149. Freynhagen R, Strojek K, Griesing T, Whalen E, Balkenohl M. (2005) Efficacy of pregabalin in neuropathic pain evaluated in a 12-week, randomised, double-blind, multicentre, placebo-controlled trial of flexible- and fixed-dose regimens. *Pain*, 115: 254-263.
 150. Jensen-Dahm C, Rowbotham MC, Reda H, Petersen KL. (2011) Effect of a single dose of pregabalin on herpes zoster pain. *Trials*, 12: 55: 1-5.
 151. Sabatowski R, Gálvez R, Cherry DA, Jacquot F, Vincent E, Maisonobe P, Versavel M, Group T-S. (2004) Pregabalin reduces pain and improves sleep and mood disturbances in patients with post-herpetic neuralgia: results of a randomised, placebo-controlled clinical trial. *Pain*, 109: 26-35.
 152. van Seventer R, Feister HA, Young JP, Stoker M, Versavel M, Rigaudy L. (2006) Efficacy and tolerability of twice-daily pregabalin for treating pain and related sleep interference in postherpetic neuralgia: a 13-week, randomized trial. *Curr. Med. Res. Opin.*, 22: 375-384.
 153. Dworkin RH, Corbin AE, Young JP, Jr., Sharma U, LaMoreaux L, Bockbrader H, Garofalo EA, Poole RM. (2003) Pregabalin for the treatment of postherpetic neuralgia: a randomized, placebo-controlled trial. *Neurology*, 60: 1274-1283.
 154. Attal N, Cruccu G, Haanpää M, Hansson P, Jensen TS, Nurmikko T, Sampaio C, Sindrup S, Wiffen P. (2006) EFNS guidelines on pharmacological treatment of neuropathic pain. *Eur. J. Neurol.*, 13: 1153-1169.
 155. Achar A, Chakraborty PP, Bisai S, Biswas A, Guharay T. (2012) Comparative study of clinical efficacy of amitriptyline and pregabalin in postherpetic neuralgia. *Acta Dermatovenerol Croat*, 20: 89-94.
 156. Zhang M, Gao C-X, Ma K-T, Li L, Dai Z-G, Wang S, Si J-Q. (2018) A Meta-Analysis of Therapeutic Efficacy and Safety of Gabapentin in the Treatment of

- Postherpetic Neuralgia from Randomized Controlled Trials. *Biomed Res Int*, 2018; 7474207: 1-10.
157. Zaccara G, Gangemi P, Perucca P, Specchio L. (2011) The adverse event profile of pregabalin: A systematic review and meta-analysis of randomized controlled trials. *Epilepsia*, 52: 826-836.
 158. Parsons B, Tive L, Huang S. (2004) Gabapentin: a pooled analysis of adverse events from three clinical trials in patients with postherpetic neuralgia. *Am. J. Geriatr. Pharmacother.*, 2: 157-162.
 159. Quintero GC. (2017) Review about gabapentin misuse, interactions, contraindications and side effects. *J. Exp. Pharmacol.*, 9: 13-21.
 160. Rullán M, Bulilete O, Leiva A, Soler A, Roca A, González-Bals MJ, Lorente P, Llobera J, Cladera M, Comas C, Mir MA, Cifre A, Lliteras B, Gestoso S, Jover A, Bestard F, Comas F, López L, Ortuño R, Peiro J, Cerdó M, Ramírez V, Gutierrez M, Argüelles R, Gutierrez MD, group PHN. (2017) Efficacy of gabapentin for prevention of postherpetic neuralgia: study protocol for a randomized controlled clinical trial. *Trials*, 18: 24: 1-9.
 161. Bulilete O, Leiva A, Rullán M, Roca A, Llobera J, on behalf of PHNG. (2019) Efficacy of gabapentin for the prevention of postherpetic neuralgia in patients with acute herpes zoster: A double blind, randomized controlled trial. *PLoS One*, 14: e0217335: 1-17.

8. Bibliography of the candidate's publications

The publications related to the PhD thesis

1. **Sadaeng W**, Marta K, Matrai P, Hegyi P, Toth B, Nemeth B, Czumbel LM, Sang-Ngoen T, Gyongyi Z, Varga G, Revesz P, Szanyi I, Karadi K, Gerber G. (2020) gamma-Aminobutyric Acid and Derivatives Reduce the Incidence of Acute Pain after Herpes Zoster - A Systematic Review and Meta-analysis. *Curr Pharm Des*, 26: 3026-3038. **IF: 3.116**
2. Sang-Ngoen T, Czumbel LM, **Sadaeng W**, Miko A, Nemeth DI, Matrai P, Hegyi P, Toth B, Csupor D, Kiss I, Szabo A, Gerber G, Varga G, Keremi B. (2021) Orally Administered Probiotics Decrease Aggregatibacter actinomycetemcomitans but Not Other Periodontal Pathogenic Bacteria Counts in the Oral Cavity: A Systematic Review and Meta-Analysis. *Front Pharmacol*, 12: 682656: 1-14. **IF: 5.811**

The publication non-related to the PhD thesis

1. Ruksakiet K, Hanak L, Farkas N, Hegyi P, **Sadaeng W**, Czumbel LM, Sang-Ngoen T, Garami A, Miko A, Varga G, Lohinai Z. (2020) Antimicrobial Efficacy of Chlorhexidine and Sodium Hypochlorite in Root Canal Disinfection: A Systematic Review and Meta-analysis of Randomized Controlled Trials. *J Endod*, 46: 1032-1041. **IF: 4.171**
2. Lukács A, Máté Z, Farkas N, Mikó A, Tenk J, Hegyi P, Németh B, Czumbel LM, **Wuttapon S**, Kiss I, Gyöngyi Z, Varga G, Rumbus Z, Szabó A. (2020) The quadrivalent HPV vaccine is protective against genital warts: a meta-analysis. *BMC Public Health*, 20: 691: 1-16. **IF: 3.295**

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